

Selection of an Anti-tau Antibody Candidate Targeting Pathological Tau for the Treatment of Alzheimer's Disease

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INTRODUCTION

The current hypothesis for the progression of tau pathology in Alzheimer's disease (AD) is that neuron-to-neuron transmission of pathologic tau, including especially trans-synaptic propagation, plays a major role. Our goal is to identify a selective, potent and efficacious anti-tau antibody clinical candidate that blocks pathologic tau spreading *in vivo* for the treatment of AD.

Immunization of mice with AD patient-derived PHF-tau (paired helical filamentous tau) as the immunogen yielded 113 anti-tau antibody hits with significant binding to PHF-tau and an absence of detectable binding to wild type recombinant tau. These antibodies were characterized and prioritized based on affinity, biophysical characteristics, efficacy in animal models of tau spreading and differentiation from clinically ineffective anti-tau antibodies.

Four anti-tau antibodies were selected (Ab01, Ab03, Ab04 and Human Ab5) with novel sequences and epitopes that fit our target profile based on selectivity, functional inhibition *in vitro* and *in vivo*, and developability. Ab01, Ab03 and Ab04 are murine antibodies that target the same C-terminal epitope, whereas Human Ab5 targets the mid-domain of tau. Among these four antibodies, Ab01 demonstrated superior efficacy in the mouse seeding model and has been humanized.

We plan to leverage the Ab01 antibody for a passive immunotherapy for AD. The clinical candidate has been chosen based on selectivity for pathological tau, potency, functional inhibition *in vitro* and developability.

OVERVIEW

Figure 1. Voyager Anti-Tau Antibody Discovery

GOAL: Identify anti-tau antibodies that block tau-seed mediated propagation of pathology

APPROACH:

- Focus on antibodies that target pathological tau species**
- Generate a diverse starting pool of anti-tau Abs**
 - Immunization Campaigns
 - Host animals: WT mouse, Tau-KO mouse for murine Ab's; human mouse for human Abs
 - Immunogens: Paired Helical Filamentous tau (ePHF or sarkosyl insoluble fraction) from AD brain
- Screen and prioritize anti-tau Abs based on:**
 - Biochemical selectivity for pathological tau
 - IHC selectivity for AD/PSP vs WT human brain
 - Functional inhibition of PHF seeding *in vitro* and *in vivo*
 - Developability based on low polyspecificity and lack of aggregation
- Characterize additional attributes**
 - Sequence
 - Epitope
 - Western blot binding to AD, PSP vs WT human brain

Mouse immunization strategy and candidates' selection profiles for identifying efficacious anti-tau candidates.

STATUS:

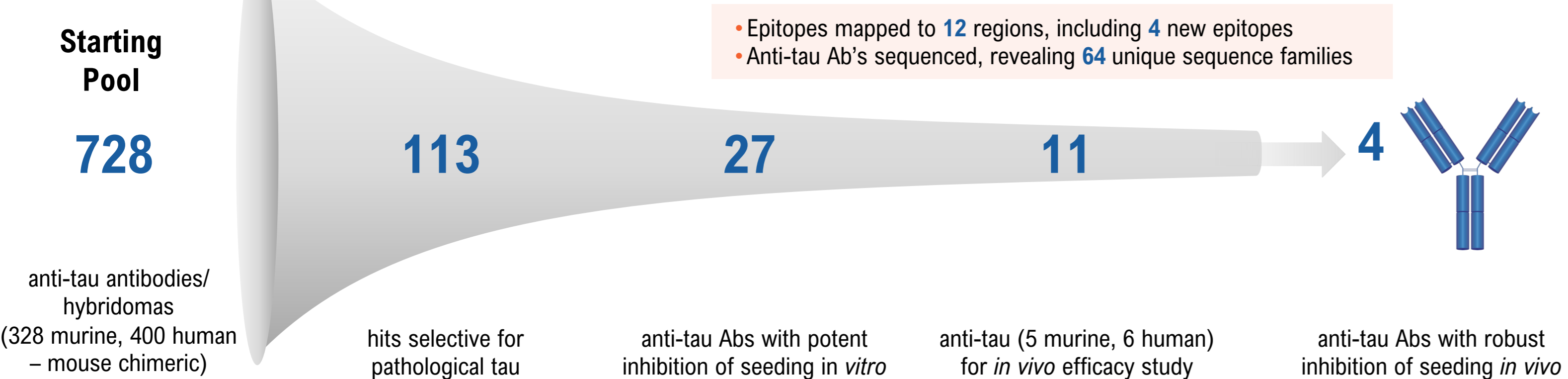
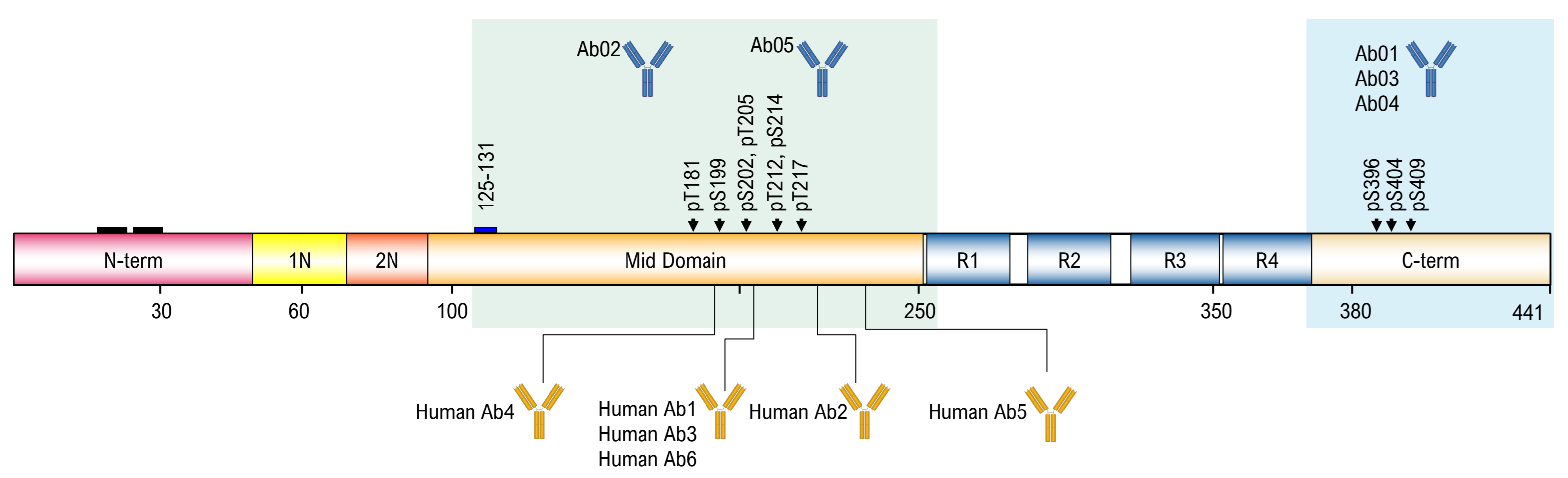


Figure 2. Voyager Anti-tau Antibodies Target Various Epitopes Throughout Full-length Tau



The 11 anti-tau Abs candidates, that were selected for *in vivo* efficacy studies, target diverse locations within full-length tau including 8 in mid-domain and 3 in C-terminus.

Table 1. Biophysical Properties of Voyager Anti-tau Antibodies

Property	Criteria*	MURINE ABS					HUMAN ABS					
		Ab01	Ab02	Ab03	Ab04	Ab05	Human Ab1	Human Ab2	Human Ab3	Human Ab4	Human Ab5	Human Ab6
Affinity (Biacore)	nM	0.044	0.3	0.058	0.12	<4.4	0.6	11	0.4	0.2	6	0.3
Selectivity: ePHF:WT rec. Tau*	≥ 100-fold	>222*	>415*	>140*	>123*	Not selective**	>23*	>59*	>40*	>137*	Not selective**	>181*
IHC Fixed - Human AD Brain	positive	Positive	Positive	Positive	Positive	Weak	Positive	Positive	Positive	Positive	Positive	Positive
IHC Fixed - Human Ctl Brain	negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative (Nu)	Negative	Negative	Negative
IHC Fixed - Human PSP	-	Positive	Positive (Wk)	Positive	Positive	Positive	Positive	Positive	Positive	Wk Positive	Positive	Positive
IHC Fixed - Human Ctl Brain	-	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative (Nu)	Negative	Negative	Negative
IHC Frozen - Human AD Brain	-	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Wk Positive	Positive	Positive
IHC Frozen - Human PSP	positive	Positive	Positive	Positive	Positive	Positive	Positive	wk Positive	Positive	Positive	Wk Positive	Positive
IHC Frozen - Human Ctl Brain	negative	Negative	Negative	Negative	Negative	weak	Negative	Negative	Negative (Nu)	Negative	Negative	Negative
Inhibition of ePHF seeding in Biosensor Cells	≤ 20 nM IC ₅₀	18.2	4.8	16.5	18.6	3.0	36.3	9.39	8.29	18.1	36	12.5
Low Polyspecificity (using BVP ELISA)	in range of comp. Abs	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
Solution and Colloidal Stability at >10 mg/mL	95% pure by SEC, no particulates	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b	✓ ^b

* No binding to highest concentration tested; ** not selective; 1:1 binding; * dash indicates not a criteria; ^a: tested at 1 mg/mL

RESULTS

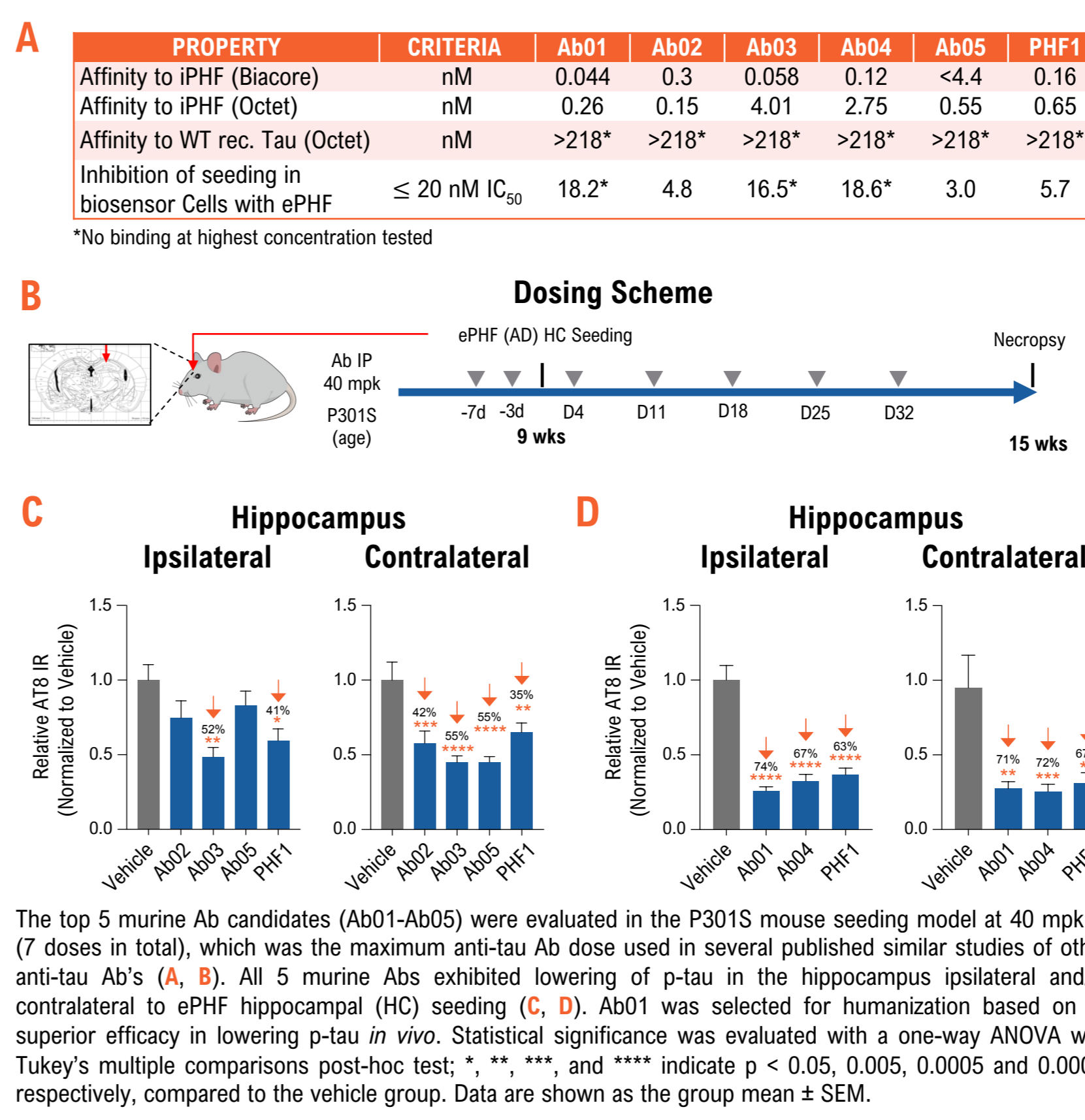
Figure 3. AD-PHF Seeding Model in P301S Transgenic Mouse

A P301S Hippocampal Seeding Model
 • FTD Mutant tau transgenic animals
 • AD-derived PHF seeding in hippocampus
 • Experiment terminated +6 weeks
 • Readout: pathological tau as assessed by AT8 ELISA (relative AT8 signal)

B Hippocampus
 Graph showing Relative AT8 Signal Normalized to AD Pathology for ePHF Inj, ePHF Con, Vehicle Inj, and Vehicle Con.

C Hippocampus, Vehicle Inj. vs Hippocampus, ePHF Inj.
 Images showing immunohistochemistry for tau pathology.

Figure 4. Reduction of p-Tau in P301S Mouse Seeding Model by Murine Antibody Candidates



The top 5 murine Ab candidates (Ab01-Ab05) were evaluated in the P301S mouse seeding model at 40 mpk IP (7 doses in total), which was the maximum anti-tau Ab dose used in several published similar studies of other anti-tau Abs (A, B). All 5 murine Abs exhibited lowering of p-tau in the hippocampus (ipsilateral and/or contralateral to ePHF hippocampal (HC) seeding (C, D)). Ab01 was selected for humanization based on its superior efficacy in lowering p-tau *in vivo*. Statistical significance was evaluated with a one-way ANOVA with Tukey's multiple comparisons post-hoc test; **, ***, **** indicate p < 0.05, 0.005, 0.0005 and 0.0001, respectively, compared to the vehicle group. Data are shown as the group mean ± SEM.

Figure 5. Affinity of Humanized Ab01 (hAb01) Variants to Immuno-purified PHF and Selectivity of hAb01 Variants for Enriched PHF vs WT Tau

A

mouse Ab01	AFFINITY TO IPHF (K _d , pM)	AFFINITY TO PTAU PEPTIDE (pM)	EPHF BINDING (EC ₅₀ , nM)	SELECTIVITY (WT 20 nM/EPHF EC ₅₀)
HC0/LC0	16.2	0.037	0.083	>133
1 HC1/LC1	29.3	38.6	0.15	>124.8
2 HC1/LC2	39.1	39.8	0.16	>149.9
4 HC2/LC1	33.4	39.8	0.133	>119
8 HC3/LC1	49.6	53.3	0.168	>232.9
9 HC3/LC2	43.4	40.8	0.085	>210.3
11 HC4/LC1	32.5	45.7	0.095	>198.4
12 HC4/LC2	44.0	76.3	0.1	>118.8
13 HC4/LC3	30.4	43.6	0.14	>113.8
15 HC5/LC1	24.9	40.3	0.164	>122
16 HC5/LC2	35.3	36.1	0.181	>128.5
17 HC5/LC3	24.8	29.4	0.191	>128.5

B hAb01 Variant Binds ePHF
 Graph showing OD450 vs hAb01 Variants nM for HC0/LC0, HC1/LC1, HC1/LC2, HC1/LC3, HC2/LC1, HC3/LC1, HC3/LC2, and HC2/LC3.

C hAb01 Variant Binds Non-Phosphorylated Recombinantly Expressed Tau
 Graph showing OD450 vs hAb01 Variants nM for IPN002 and All 18 variants.

Eleven of 18 humanized Ab01 variants were the focus of further work based on selectivity and affinity. A) iPHF and pTau peptide affinity measurements by Biacore. Binding affinity was measured using Surface Plasmon Resonance (SPR) on Biacore 8K instrument. For iPHF binding, iPHF was directly immobilized on CMS sensor chip by amine coupling at 199 RU density. For pTau binding, 1 µg/ml biotinylated pTau peptide was captured on Biotin CAP chip via CAPture reagent to achieve 5-10 RU levels. Antibody was injected using Single Cycle Kinetics (SCK) mode with association and dissociation times of 5 and 10 min, respectively at a concentration range of 0.78 to 2.5 nM. The sensorgrams were fitted to 1:1 binding model in the Biacore Evaluation software to determine kinetic rate constants and affinity values. HC0/LC0: mouse variable/human constant chimeric served as a control (red). B, C) Examples of affinities measured by ePHF or WT tau ELISA. Serial 3- fold dilutions of each antibody variant were prepared starting from 66 nM for a total of 12 antibody dilutions. These dilutions were exposed to a plate coated with ePHF or WT tau for a direct ELISA as described in Liu, et al (2016). For each variant, OD450 readings were plotted against the corresponding antibody concentration. The EC₅₀ was then determined by non-linear regression using Graphpad Prism.

Figure 6. All Top 11 Humanized Ab01 Variants Selectively Immunostain Tangles in AD Cortex

Grid of immunohistochemistry images showing selective staining of tangles in AD cortex for variants HC4/LC1, HC1/LC2, HC2/LC1, HC3/LC1, HC3/LC2, HC4/LC2, HC4/LC3, HC5/LC1, HC5/LC3, HC5/LC2, and HC0/LC0*.

All 11 hAb01 variants demonstrated specific binding to neuronal tau pathology (white arrows) with minimal staining of non-AD cortex. *HC0/LC0: mouse variable/human constant chimeric control. 20X magnification.

Table 3. Summary of *in Vitro*, Biochemical and Biophysical Characteristics for Top 11 Humanized Ab01 Variants

PROPERTY	mAb01	HC0/LC0	HC1/LC1	HC1/LC2	HC2/LC1	HC3/LC1	HC3/LC2	HC4/LC1	HC4/LC2	HC4/LC3	HC5/LC1	HC5/LC2	HC5/LC3
Affinity to iPHF (Biacore, pM)	16.5	16.2	29.3	39.1	33.4	49.6	43.4	32.5	44	30.4	24.9	35.3	24.8
Affinity to pS422 tau peptide (pM)			38.6	39.8	39.8	53.3	40.8	45.7	76.3	43.6	40.3	36.1	29.4
Affinity to ePHF EC ₅₀ (nM)		0.08	0.15	0.16	0.13	0.17	0.085	0.095	0.1	0.14	0.16	0.18	0.19
Selectivity: ePHF:WT rec. Tau ^a		>240	>133	>124.8	>149.9	>119	>232.9	>210.3	>198.4	>118.8	>113.8	>122	>128.5
IHC Fixed - Human AD Brain		Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive
IHC Fixed - Human Ctl Brain		Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative
Western Frozen - Human AD Brain		Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive	Positive
Western Frozen - Human Ctl Brain		Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative	Negative
Inhibition of ePHF seeding in Biosensor Cells	18.2		78.1	17.6	49.8	17.6	32.2	58.1	49.8	34.5	24.1	37.9	43.8
Low Polyspecificity (using BVP ELISA)		3.9	1.8	1.6	6	1.6	1.7	2.2	1.8	3.2	1.6	1.4	2.7
PTM lability			0.505	0.505	0.505	0.505	0.505	0.505	0.505	0.505	0.505	0.505	0.505
Predict aggregation		Inc.	Low	Inc.	Inc.	Inc.	Inc.	Inc.	Inc.	Low	Low	Low	Low

^a: no binding at highest concentration tested; Inc.: inconclusive; gold boxes indicate top 5 humanized Ab01 variants selected for further developability work

Figure 7. T-Cell Immunogenicity Assay (CD4+ Cell Proliferation)

SI Distribution
 Graph showing Stimulation Index for Herceptin, HC1/LC1, HC3/LC2, HC4/LC1, HC5/LC2, and HC5/LC3.

Table of Antigen Immunogenicity:

ANTIGEN	# OF DONORS	% OF DONORS
KLH	50	100
Herceptin	4	8
Voyager_1 (HC1/LC1)	27	54
Voyager_9 (HC3/LC2)	12	24
Voyager_11 (HC4/LC1)	23	46
Voyager_16 (HC5/LC2)	24	48
Voyager_17 (HC5/LC3)	25	50

Relative immunogenicity risk for 5 humanized Ab01 variants was assessed by the CD4+ T-cell proliferation assay. Antibodies were incubated with PBMC cells from 50 healthy donors representative of the global population based on HLA-DRB1 expression and cell proliferation was measured by flow cytometry. Two assay readouts: 1. Stimulation Index (SI): ratio of # of proliferating T cells of sample over blank. SI ≥ 2.0 is considered positive response. 2. Percentage of responding donors.

CONCLUSIONS

- Diverse starting pool of proprietary Abs targeting pathological tau from human AD brain were generated and screened
- Four Abs (Ab01, Ab03, Ab04 and Human Ab5) with novel sequences and epitopes in the mid-domain or C-terminus of tau, were selected that fit the target profile based on selectivity, functional inhibition *in vitro* and *in vivo*, and developability
- Ab01 demonstrated superior efficacy *in vivo* in the P301S mouse seeding model in reducing pathological tau, and was chosen for humanization
- Humanization of Ab01 has been completed and Voyager_9 (HC3/LC2) has been selected as the clinical candidate VY-TAU01